

Instructions for Use (Handbook) MagPurix® Plant DNA Extraction Kit

Catalog No.: ZP02014

Manual No.: IFU-MP02-02014

Version: 2.0







ZINEXTS LIFE SCIENCE CORP.

16F., No. 93, Sec. 1, Xintai 5th Rd., Xizhi Dist., New Taipei City 221416, Taiwan (R.O.C.)

Read and follow these Instructions for Use prior to using this product. The latest revision of this document can be found at www.zinexts.com

Zinexts Life Science Corp. www.zinexts.com

Contents

Intended Use	2
Introduction	
Kit Contents and Storage	3
Materials Required but not Provided	4
Warnings and Precautions	4
Purification Principle	5
Things to Do Before Starting	6
Sample Preparation	6
Procedure of MagPurix System	8
Purification Protocol - MagPurix® series	8
Purification Protocol - MagPurix® EVO series	9
Purification Protocol - MagPurix® N.E.O	10
Troubleshooting	12
Related Products	13
References	13
Limited Product Warranty	13
Revision History	14

Intended Use

The MagPurix® Plant DNA Extraction Kit provides a complete set of reagents and consumables for the automated purification of genomic DNA from plant tissue and yeast using the MagPurix system.

The product is intended to be used by professional users, such as technicians and physicians who are trained in molecular biology techniques.

For research use only. Not for use in diagnostic procedures.

Introduction

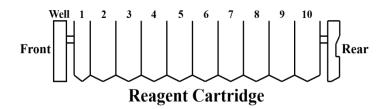
Product Name	MagPurix® Plant DNA Extraction Kit
Catalogue Number	ZP02014
Product Overview	The MagPurix® Plant DNA Extraction Kit is designed to extract genomic DNA from plant tissue and yeast using MagPurix® series automatic instruments. The kit is applied with unique magnetic ZiBeads® technology, which achieves superior product quality, consistent and high product yield and reproducible results. The purified DNA is suitable for a wide range of diagnostic and research applications, including sequencing, genotyping and qPCR detection.
Applicable	All MagPurix® Instruments
Instruments Model	
Display Protocol	2014 PLANT DNA (For MagPurix® 12/24, EVO)
Name on the	
Instrument	
Applicable	Please check and download the latest firmware from
Instrument	www.zinexts.com
Firmware	

Kit Contents and Storage

onitonito ana o	tolago	
Shipping and Storage	The kit is shipped at room temperature.	
	Upon receipt, store the kit at room temperature.	
	All kit components are stable when stored properly ur	ntil the
	expiration date shown on the kit box.	
Kit Content	The components supplied in the kit are listed below.	
	Sufficient reagents are supplied to perform 48 purifica	ations.
	Contents	Amount
	1 Reagent Cartridge	8 pcs
	Reaction Chamber (For MagPurix® 12/24, EVO)	8 pcs
	Tip Holder (For MagPurix® 12/24, EVO)	8 pcs
	4 Piercing Pin	50 pcs
	5 Filter Tip	50 pcs
	6 Sample Tube (2 ml)	50 pcs
	Z Elution Tube (1.5 ml)	50 pcs
	Process Rack (For MagPurix® N.E.O. only)	48 pcs
	RNase A, 10 mg/mL (0.5 ml)	1 pc
	PLA Buffer (25 ml)	1 pc
	PLB Buffer (25 ml)	1 pc
	Barcode Sticker (For MagPurix® EVO, N.E.O.)	50 pcs

Reagent Cartridge Contents Each Reagent Cartridge has 10 positions with 10 sealed wells. Positions 1-10 contain wells filled reagents for this protocol.

Reagent	Well No.
Empty	1
Lysis Buffer 2	2
Binding Buffer 1	3
Magnetic Bead Solution	4
Washing Buffer 1B	5
Washing Buffer A	6
Washing Buffer B	7
Elution Buffer 1	8
Elution Buffer 2	9
Empty	10



Materials Required but not Provided

The following general laboratory equipment and consumables are required to perform the extraction. All laboratory equipment should be installed, calibrated, operated, and maintained according to the manufacturer's recommendations. The following table lists the required equipment and consumables.

For all purification procedures:

- 1. MagPurix® / MagPurix® EVO series / MagPurix® N.E.O. instrument
- 2. 1.5 or 2.0 ml microcentrifuge tubes
- Pipettes and filter tips
- 4. Phosphate-buffered saline (PBS, may be required for diluting samples)
- 5. **Optional**: Plastic consumables, DNase-free RNase A (to minimize RNA content)

Warnings and Precautions

For research use only. Read all the instructions carefully before using the kit. Use of this product should be limited to trained personnel in the techniques of DNA purification. Strict compliance with the user manual is required for optimal results. Attention should be paid to expiration dates printed on the box and labels of all components. Do not use a kit after its expiration date.

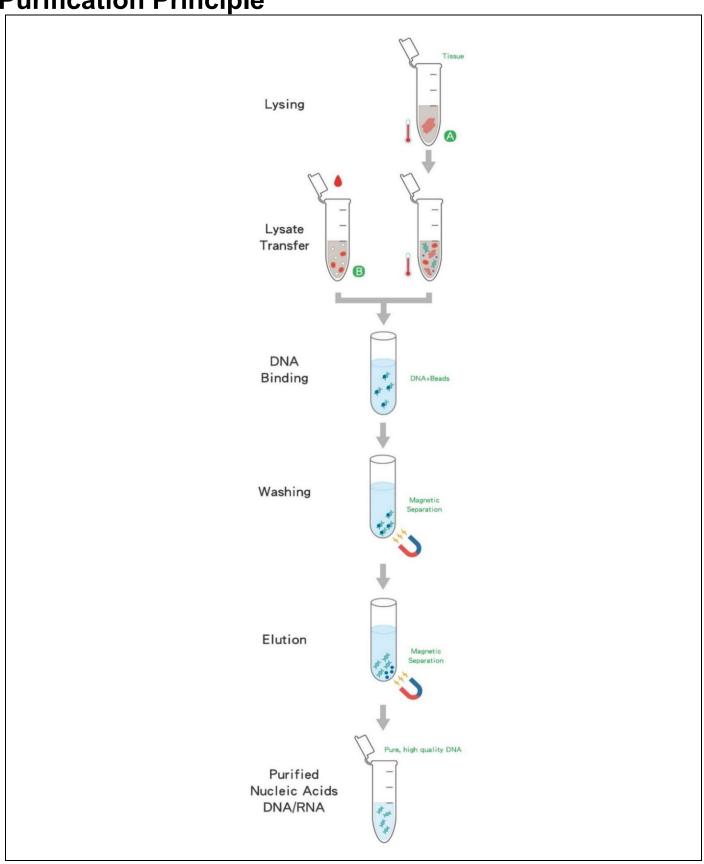
When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, please consult the appropriate safety data sheets (SDSs). These are available online in convenient and compact PDF format at MSDS (Material Safety Data Sheets) – Downloads – www.zinexts.com.

Please report any serious incident occurred in relation to the device to your local representative/ agent or the manufacturer, and to the competent authority of your country/state.



CAUTION: DO NOT add bleach or acidic solutions directly to the sample preparation waste.

Purification Principle



- Perform certain pretreatment process before extraction.
- B Transfer sample to extraction directly.

Things to Do Before Starting

Sample Preparation

The purification procedure is optimized for the use of appropriate samples as listed in table below.

Plant tissue a. Perform homogenization by using proper homogenizer. b. Add 440 µl Lysis Buffer to the sample. c. Vortex vigorously. d. Incubate the mixture at 65°C for 10 minutes in a thermomixer (set 1000 rpm) or vortex several times during incubation in the heat block water bath. e. Pre-filter the digested lysate using a Filter Column to remove residudebris. f. Short spin at 6.000 x g to collect the clear flow-through in Collection	or ual ion
 c. Vortex vigorously. d. Incubate the mixture at 65°C for 10 minutes in a thermomixer (set 1000 rpm) or vortex several times during incubation in the heat block water bath. e. Pre-filter the digested lysate using a Filter Column to remove residudebris. f. Short spin at 6.000 x g to collect the clear flow-through in Collection 	or ual ion
 d. Incubate the mixture at 65°C for 10 minutes in a thermomixer (set 1000 rpm) or vortex several times during incubation in the heat block water bath. e. Pre-filter the digested lysate using a Filter Column to remove residudebris. f. Short spin at 6.000 x g to collect the clear flow-through in Collection 	or ual ion
 1000 rpm) or vortex several times during incubation in the heat block water bath. e. Pre-filter the digested lysate using a Filter Column to remove residudebris. f. Short spin at 6.000 x g to collect the clear flow-through in Collection 	or ual ion
 1000 rpm) or vortex several times during incubation in the heat block water bath. e. Pre-filter the digested lysate using a Filter Column to remove residudebris. f. Short spin at 6.000 x g to collect the clear flow-through in Collection 	or ual ion
debris. f. Short spin at $6.000 \times g$ to collect the clear flow-through in Collecting	ion
· · · · · · · · · · · · · · · · · · ·	
· · · · · · · · · · · · · · · · · · ·	
Tube.	om
g. Add 10 μl RNase A, mix well, incubate for 10 minutes at roo	
temperature.	
h. Transfer 100-400 μl into each Sample Tube.	
Yeast/suspension a. Centrifuge at 6.000 x g, 3 minutes.	
culture b. Remove supernatant.	
c. Add 440 µl Lysis Buffer, vortex for 30 seconds.	
d. Incubate the mixture at 65°C for 10 minutes in a thermomixer (set	at
1000 rpm or vortex several times during incubation in the heat block water bath.	or
e. Pre-filter the digested lysate using a Filter Column to remove residudebris.	ual
f. Short spin at $6.000 \times g$ to collect the clear flow-through in Collecti	ion
Tube.	.011
g. Transfer 100-400 μl into each the Sample Tube.	
Yeast/colony a. Take 1-3 colony from culture plate with an inoculation loop and suspe	nd
in 440 μl of Lysis Buffer by stirring vigorously.	
b. Incubate the mixture at 65°C for 10 minutes in a thermomixer (set	at
1000 rpm) or vortex several times during incubation in the heat block	cor
water bath.	
 c. Pre-filter the digested lysate using a Filter Column to remove residu 	ual
debris.	
d. Short spin at $6.000 \times g$ to collect the clear flow-through in Collecti	ion
Tube.	
e. Transfer 100-400 μl suspension into each Sample Tube.	

Note:

After harvesting plant tissues, it should be frozen in liquid nitrogen if not be used immediately. It can then be stored at -80°C. Alternatively, tissue can be dried or lyophilized after harvesting to allow storage at room temperature (15-25°C). To ensure DNA quality, samples should be completely dried within 24 hours of collection. If possible, it is preferable to collect young materials (e.g., leaves, needles) since they contain more cells per weight and therefore result in higher yields.

When working with fungi, harvest mycelium directly from a culture dish or from liquid culture. For liquid culture, first pellet cells by centrifugation. Remove the supernatant completely before disruption and lysis. Fresh, frozen, or freeze-dried fungal material can be used.

The disruption method may require optimization to ensure maximum DNA yield and quality. Complete and quick disruption of starting material is essential to ensure high DNA yields and to avoid DNA degradation.

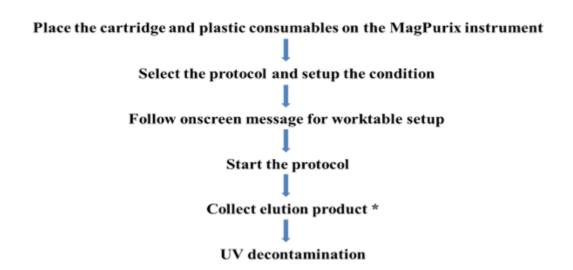
Before DNA extraction, plant material should be first mechanically disrupted with Lysis Buffer. After homogenization, remove the debris and other precipitations by using Filter Column. Collect the clear flow-through and incubate with RNase A to digest the RNA in the sample before DNA extraction.

We provide two Lysis Buffers: **PLA Buffer** and **PLB Buffer** for dealing with different tissue types. Before extraction of a new tissue type, try both the Lysis Buffers to get the optimized lysis procedure and a better DNA yield. If the precipitation formed in the Lysis Buffer, warm it at 65 °C before use.

Table A – The sugg extraction	ested starting material and e	elution volume range for each n	ucleic acid		
Sample type	Starting material per sample	Concentration (mg/µl)	Elution Volume		
Plant tissue					
Soybean	100-400 µl/100 mg seed	5-12 (PLA)/50-80 (PLB)			
Rice	100-400 µl/20 mg seed	5-8 (PLA)/15-25 (PLB)			
Arabidopsis		2-5 (PLA)/5-7 (PLB)			
Tomato		20-40 (PLA)	50-300 μΙ (EVO 50-200 μΙ)		
Corn		10-15 (PLA)/25-60 (PLB)			
Tectaria	100-400 μl/100 mg leaf	5-10 (PLA)			
Aspidistra		3-6 (PLA)			
Pharius		20-25 (PLA)/50-100 (PLB)			
Zingiber		3-8 (PLA)/20 -25 (PLB)			
Yeast					
Suspension Cultures	100-400 μl		50-300 μl (EVO 50-200 μl)		
Colony	1-3 colony		(L v O 30-200 μι)		

Procedure of MagPurix System

Workflow of MagPurix operation



^{*} Download the run record (MagPurix EVO & MagPurix N.E.O. series)

Purification Protocol - MagPurix® series

1	Turn on the Instrument	a.	Turn on the power switch and wait for the screen to turn on.
7	Load new	a.	Open the door and remove the Sample Rack from the instrument.
_	Consumable(s)	b.	Load 1 Reagent Cartridge, and all plastic disposables (2 Reaction
	and Cartridge(s)		Chamber, 3 Tip Holder, 4 Piercing Pins, 5 Filter Tips and other components presented in the kit intended to use).
		C.	Place 6 Sample Tubes and 7 Elution Tubes into the Sample Rack.
3	Load the	a.	Transfer appropriate volume of sample into each Sample Tube on the
O	Samples		Sample Rack.
			Put the Sample Rack back into the instrument and close the door.
4	Program Set up	a.	Scan the protocol barcodes to select the purification protocol, sample volume and elution volume.
5	Start Extraction	a.	Follow the instructions displayed on the screen to double-check the operating steps being completed before program running.
		b.	Press " ENTER " to start the experiment. Instrument will run the protocol program automatically until the whole process is completed.
		C.	At the end of the run (approximately 12 series 45-60 minutes, 24 series 50-65 minutes), instrument alarms briefly.
6	Collect the	a.	
O	Elution Tubes	b.	Collect the Elution Tubes containing the purified nucleic acids.
		C.	

analysis.

- d. Discard the used cartridges and all plastic consumables into biohazard waste. *Do not reuse the cartridges.
- e. If you are not using the instrument immediately, place the Sample Rack back to the workplace, close the instrument door and press "Start" button for 2 seconds to enter sleep mode. If the instrument will not be used in an extended period of time, please turn off the power switch.

Purification Protocol - MagPurix® EVO series

1	Turn on the	a.	Turn on the power switch and wait for the screen to turn on.
ı	Instrument	a.	Login the instrument and enter the Home Page.
2	Load new	a.	Open the door and remove the Sample Rack from the instrument.
_	Consumable(s)	b.	Open the Tip-Holder Lid.
	and Cartridge(s)	C.	Load IReagent Cartridge and all plastic disposables (2 Reaction
			Chamber, 3 Tip Holder, 4 Piercing Pins, 5 Filter Tips and other
			components presented in the kit intended to use).
		d.	Close the Tip-Holder Lid.
		e.	Paste the Barcode Stickers on Elution Tubes.
		a.	Place 6 Sample Tubes and 7 Elution Tubes into the Sample
			Rack.
2	Load the	a.	Transfer appropriate volume of sample into each Sample Tube on the
J	Samples		Sample Rack.
		a.	Put the Sample Rack back into the instrument and close the door.
1	Program Set up	a.	Select the appropriate protocol program on the instrument. Press
_		_	NEXT.
		b.	Select the appropriate Sample Volume and Elution Volume and press
			NEXT.
		C.	Press the number button to select the right Sample Numbers.
		d.	Scan/Edit each primary Sample ID directly. After finished, press NEXT .
		•	Scan/Edit each Elution Tube ID directly. After finished, press NEXT .
		e. f.	Scan Reagent Cartridge Barcode. Press NEXT .
		١.	*If the cartridge is expired, the next step cannot be performed.
		a.	Follow the instructions on the screen to double-check the operating
		u.	steps being completed before running the program. Press NEXT .
	Start Extraction	a.	Check "PROGRAM CONFIRMATION" on the screen.
5		b.	Press "START" to start the experiment. Instrument will run the
			protocol program automatically until the whole process is completed.
		C.	At the end of the run (approximately 40-45 minutes), instrument
			alarms briefly and the screen indicates "PROGRAM FINISH".
		d.	If you want to perform the same protocol, press "RERUN" to
			perform the same experiment. If you do not need to re-run the
			experiment, press the function button " HOME" to exit the
			experiment mode.
6	Collect the	a.	Open the instrument door.
O	Elution Tubes	b.	Collect the Elution Tubes containing the purified nucleic acids.
		C.	The purified nucleic acids are ready for immediate use. Store the

- purified nucleic acids at 4°C (short-term, less than 10 days) or aliquot and store at -70°C (long-term) before performing downstream analysis.
- d. Discard the used cartridges and all plastic consumables into biohazard waste. *Do not reuse the cartridges.
- e. If you are not using the instrument immediately, please put the Sample Rack back into the instrument, close the instrument door, and press the "POWER" function button to enter sleep mode. If the instrument will not be used in an extended period of time, please turn off the power switch.

Purification Protocol - MagPurix® N.E.O.

			•
1	Turn on the Instrument	a. b.	Turn on the power switch and wait for the screen to turn on. Scan the user personal barcode to Login the instrument and enter the Home Page.
2	Program set up	a. b. c. d.	Scan the barcode of the MagPurix® Extraction kit. For optimum results, always use a kit within the expiry time mentioned on the kit box. Use the +/- buttons or manually enter the input total volume of sample after facultative pretreatment and the elution volume required. Press . ATTENTION: the drawer will open immediately, keep clear from the drawer opening area. Look at the 2 pop-up animations windows that teach how to 1-Select sample position, 2- scan sample IDs, press "NEXT". Select whether your samples belong to a "working list". If yes, MagPurix® N.E.O. will recognize the samples by connecting to your organization LIS network. Select a sample position between 1-12, scan all sample tube barcodes and elution tube barcodes. Press when all samples
			are edited.
3	Load new Consumable(s) and Cartridge(s)	a.	Verify that all samples are all set properly, place onto the worktable all consumables, Reagent Cartridge and all plastic disposables (4 Piercing Pins, 5 Filter Tips, 8 Process Rack and other components presented in the kit intended to use).
4	Load the Samples	a. b.	Transfer appropriate volume of sample into each Sample Tube on the Sample Rack. Place the Sample Tubes and Elution Tubes on the MagPurix® N.E.O. Sample Rack, following the same order as set on the MagPurix® N.E.O. system. Press "Close drawer", the drawer will close automatically.
5	Start Extraction	a.	Press after the drawer has closed NOTE: It is possible to Pause to resume or to abort the extraction process. Press to resume or to abort the extraction process.
6	Collect the Elution Tubes	a. b.	The Extraction process is finalized (approximately 40-45 minutes) when alarm rang and the MagPurix [®] N.E.O. will display the extraction process report. Press "Export" to export the Data report to an USB drive. Data reports are stored in Toolbox>data archive

- c. Press to terminate the experiment. ATTENTION: the drawer will open immediately, keep clear from the drawer opening area.
- d. Collect the Elution Tubes containing the purified nucleic acids.
- e. The purified nucleic acids are ready for immediate use. Store the purified nucleic acids at 4°C (short-term, less than 10 days) or aliquot and store at -70°C (long-term) before performing downstream analysis.
- f. Discard the used cartridges and all plastic consumables into biohazard waste. *Do not reuse the cartridges.
- g. Press "Close drawer" then , the MagPurix® N.E.O. system will automatically redirect to the UV decontamination page.
- h. Press "**UV Decontamination**", and select the desired time using +/- buttons. Press "**Start**".
 - **NOTE:** It is possible to Pause the decontamination process. Press to resume or to abort the decontamination process.
- Press "OK" when the decontamination process is finished. MagPurix[®] N.E.O. will redirect to the LOGIN page.

Troubleshooting

*This table is helpful for solving common problem. If you need other technical support, please contact Zinexts team (sales@zinexts.com) or your distributor.

Problem	Possible Cause	Comments and suggestions
Poor DNA quality or yield	Deterioration or contamination of reagents.	Please ensure that the reagents of kit are still within the effective shelf-life period before use. Discard any kit reagent that shows discoloration or evidence of microbial contamination.
	Kit stored under non-optimal conditions.	Store kit at 15-25°C at all times after arrival. If either Reagent or Buffer precipitates upon shipping in cold weather or during long-term storage, dissolve precipitates by gently warming and stirring the solution. Please do not freeze the Reagent Cartridges.
	Insufficient sample input.	DNA yield depends on the sample type and the number of nucleated cells in the sample. Please proportionally adjust the total input amount of sample to increase the DNA yield.
	Too much of elution buffer was used.	The elution volume can be reduced proportionally.
	The eluent of final product(s) is not enough.	Please collect issue information and provide it to your Support Representative /Technical Support as soon as possible.
Clogged issue	Too much sample material was used.	Decrease the input amount of sample material or dilute your sample.
No results in downstream analysis	No signal/The PCR was inhibited.	Using appropriate controls for analysis. Check the positive control, negative control, water (NTC) and internal control to clarify the possible causes.
Instrument malfunction/abnormal sound	Abnormal consumables: 1. Deformed Filter Tips 2. Deformed Reaction Chamber 3. Deformed Tip Holder	Please replace the batch with normal consumables.
	Abnormal action of instrument: 1. Inaccurate correction value 2. Spare parts or components damaged	Please collect issue information (videos and pictures) and provide it to your Support Representative/Technical Support as soon as possible to calibrate or replace any other damaged or worn parts.

Related Products

Product Name	Cat. No.
MagPurix® Blood DNA Extraction Kit 200	ZP02001
MagPurix® Blood DNA Extraction Kit 1200	ZP02002
MagPurix® Viral Nucleic Acid Extraction Kit	ZP02003
MagPurix® Tissue DNA Extraction Kit	ZP02004
MagPurix® Cultured Cell DNA Extraction Kit	ZP02005
MagPurix® Bacterial DNA Extraction Kit	ZP02006
MagPurix® HPV DNA Extraction Kit for Swab Samples	ZP02007
MagPurix® TB DNA Extraction Kit	ZP02008
MagPurix® FFPE DNA Extraction Kit	ZP02009
MagPurix® Forensic DNA Extraction Kit	ZP02010
MagPurix® Viral/Pathogen Nucleic Acids Extraction Kit A	ZP02011
MagPurix® Viral/Pathogen Nucleic Acids Extraction Kit B	ZP02012
MagPurix® Viral RNA Extraction Kit	ZP02013
MagPurix® Plant DNA Extraction Kit	ZP02014
MagPurix® Total RNA Extraction Kit	ZP02015
MagPurix® Viral Nucleic Acid Extraction Kit LV	ZP02016
MagPurix® CFC DNA Extraction Kit	ZP02017
MagPurix® Coronavirus RNA Extraction Kit	ZP02027
MagPurix® Urine cfDNA Extraction Kit	ZP02032
MagPurix® Plasma cfDNA Extraction Kit	ZP02033

References

• Tan SC et al. J Biomed Biotechnol. (2009)

Limited Product Warranty

Zinexts Life Science Corp. is committed to provide customers with high-quality products and services. Our goal is to ensure that every customer is 100% satisfied with our products and services. If you have any question or concerns, contact our Technical Support Representatives.

Zinexts Life Science Corp. guarantees the performance of all products according to the specifications stated in our product literature. The purchasers/users must determine the suitability of the product for their particular use. We reserve the right to change, alter, or modify any product to enhance its performance and design.

This warranty limits the liability of Zinexts Life Science Corp. to only the cost of the product. No warranty is granted for products beyond their listed expiration date. No warranty is applicable unless all product components are stored and used in accordance with instructions.

Revision History

Version	Date	Description
2.0	26. Sept. 2024	1. Add application for MagPurix® N.E.O.
		2. Revise related products
		3. Update symbols